

Chapter 9

Bioabsorbable Polymer and Bone Growth Factor Composites

L.-M. Tielinen

Summary

Bone growth factors are small proteins contained by bone tissue. The most intensively investigated bone growth factors are the bone morphogenetic proteins (BMPs) and the transforming growth factor- β (TGF- β s). Their local application induces bone formation in experimental fractures and bone defects. Possible clinical applications for bone growth factors include impaired bone healing, inadequate bone stock, and incomplete incorporation of prosthetic implants into bone. In therapeutic use the bone growth factors need a carrier. The carrier should assure local, sustained release of the growth factors which may otherwise be rapidly absorbed before instituting their effect. Several biodegradable materials have been investigated as carriers for bone growth factors, including organic materials, ceramics, and synthetic polymers such as polylactide and polylactide-polyglycolide co-polymer. Bioabsorbable polymers possess several attractive features as delivery systems. They can be engineered to allow release of accurate dosage of bone growth factors and they also allow combining several growth factors to be delivered concomitantly or at different points of time. They can also be manufactured or sculpted into a variety of forms to fit a given tissue defect or to fix a fracture. Moreover, they are both sterilizable and can be resorbed completely. An increasing number of positive results from experimental studies exist which could indicate a possible future success for growth factors within clinical orthopaedic practice. Recently, the first human studies using recombinant human BMP-2 or BMP-7 (osteogenic protein-1, OP-1) have been published.

*Correspondence to: L.-M. Tielinen, Töölön sairaala, PL 266, 00029 HUS, Helsinki, Finland. E-mail: laura.tielinen@hus.fi

Bone growth factors are small proteins contained by bone tissue. The most intensively investigated bone growth factors are the bone morphogenetic proteins (BMPs) and the transforming growth factor- β s (TGF- β s). Their local application induces bone formation in experimental fractures (1,2) and bone defects (3,4). A simplistic summary may be that TGF- β , released by platelets at the fracture haematoma, stimulates cells to make and to react to BMPs and other growth factors in a synergistic cascade. This cascade, given the right mechanical conditions and the presence of new blood supply, leads to regenerative bone repair.

Possible clinical applications for bone growth factors include impaired bone healing, inadequate bone stock, and incomplete incorporation of prosthetic implants into bone. Complicated fractures with extensive soft tissue damage often heal insufficiently, resulting in delayed union or nonunion. Large amounts of bone grafts are needed in bone tumour surgery, endoprosthetic surgery, and reconstructive and maxillofacial surgery. Today, the above-mentioned clinical problems are solved by bone grafting. Autogeneic bone graft from the iliac crest is the main source of trabecular bone. It has a good osteoinductive capacity, but the sources are limited and the harvest procedure causes postoperative discomfort to the patient. Allogeneic bone is also widely used but provides mainly osteoconductive properties. Moreover, despite of extensive testing, there are still potential risks for transmitting diseases.

Bone-derived growth factors are produced by osteoblasts and incorporated into the extracellular matrix during bone formation. Trauma or remodeling causes solubilization of the proteins (5). After release, the growth factors may initiate and control a healing response after bone trauma and they are able to regulate osteoblast and osteoclast metabolism during bone remodeling. They exhibit their effects only in the local environment. Most growth factors are released as high molecular weight precursors which are split by proteolysis to produce active factors which are generally of a low molecular weight (6). They exhibit their effects by binding to membrane bound receptors on the surface of the target cells. This leads to a cascade of intracellular events which affect the expression of genes that encode for such functions as protein synthesis and cell division. The total number of identified growth factors able to affect proliferation, differentiation, and secretive functions of bone-related cells is continually expanding as a result of research.

The following bone growth factors are the most important in the bone tissue. Transforming growth factor-beta (TGF- β) exists in five different subtypes (TGF β 1-5). TGF- β is probably the most potent multifunctional regulator of bone cell metabolism. Fifteen bone morphogenetic proteins (BMPs) have, so far, been identified (7). They are the only growth factors known to stimulate the mesenchymal stem cells to differentiate into osteoblastic and chondroblastic lineage. Platelet-derived growth factors (PDGFs) exist in three isotypes and are released from the platelets at the site of injury. They are potent mitogens for cells of connective tissue origin and are thought to play a role in soft tissue and fracture healing (8). Insulin-like growth factors (IGF I and II) are produced by osteoblasts. They have been suggested to have important functions for skeletal growth by mediating the growth-promoting actions of growth hormone (9). Fibroblast growth factors (FGFs) are secreted by osteoblasts. The prototype members of the fibroblast growth factor family are the acidic fibroblast growth factor (aFGF, FGF-1) and the basic fibroblast growth factor (bFGF, FGF-2) (10). They are best known for their effects on endothelial cell replication and neovascularization (11). PDGFs, IGFs, and FGFs are all mitogenic factors produced in early phases of fracture repair and localized in the bone matrix (12). However, their true therapeutic potential for fracture healing has not yet been revealed.

In therapeutic use the bone growth factors need a carrier. The carrier should assure local, sustained release of the growth factors which may otherwise be rapidly absorbed before instituting their effect.

Several biodegradable materials have been investigated as carriers for bone growth factors, including (1) organic materials such as inactive demineralized bone proteins (13,14), collagen (15,16), fibrin (17,18), squalene (19,20), and coral (21,22); (2) ceramics, including tricalcium phosphate (23,24), hydroxyapatite (25,26), calcium-sulphate composites (27,28), and bioactive glasses (29,30); and (3) synthetic polymers such as polylactide (31,32,33), polylactide-polyglycolide co-polymer (34,35,36), polyanhydride (37), and polyorthoester (38,39).

Bioabsorbable polymers possess several attractive features as delivery systems. They can be engineered to allow release of accurate dosage of bone growth factors and they also allow combining several growth factors to be delivered concomitantly or at different points of time. They can also be manufactured or sculpted into a variety of forms to fit a given tissue defect or to fix a fracture.

Moreover, they are both sterilizable and can be resorbed completely. The disadvantages of the bioabsorbable polymers include the foreign body reactions due to the degradation products (40,41,42,43). The porosity of the polymer material is often insufficient but can be increased by developing new materials or combining them to porous materials, such as demineralized bone matrix, to improve their osteoconductive properties. Polylactide (44,45,46,47,48,49) and the copolymer of polylactide and polyglycolide (50,13,51,52,53) are the most frequently used components of bioabsorbable carriers for bone growth factors in experimental studies.

Polyglycolide (PGA) and polylactide (PLA) are members of a large family of poly- α -hydroxy acids. These polymers can be manufactured by polymer processing methods into various shapes such as films, rods, plates and screws. Polyglycolide (PGA) or polyglycolic acid can be synthesized from glycolide (cyclic diester of glycolic acid l. hydroxyacetic acid) under the influence of an inorganic metal salt catalyst at low concentrations by a ring-opening polymerization. Polylactide (PLA) or polylactic acid is a synthetic polymer of lactide (cyclic diester of lactic acid). It can also be synthesized by the direct condensation polymerization of the monomer lactic acid. Polylactic acid has two enantiomeric forms, optically active stereoisomers, poly-L-lactic acid (PLLA) and poly-D-lactic acid (PDLA), with similar intrinsic chemical properties but opposite conformational structures (54). The physical properties of the co-polymers of L-lactic acid and D-lactic acid (PDLLA) are dependent on the relative amounts of L-and D-monomers in the polymer chain. The degradation process of PGA and PLA begins with molecular degradation; random hydrolysis starts in aqueous environment. The high surface area/weight ratio, porosity of the implant, low initial M_w , and high monomer concentration all speed up the molecular degradation process (54,55,56). The events are followed by a decrease of strength and mass of the implant, microscopic degradation in tissue, and transfer of the monomer lactic acid or glycolic acid to the metabolism of cells (57,58).

Our research group used a blend of an L-lactic acid oligomer and a co-polymer of epsilon caprolactone and DL-lactic acid as a carrier for TGF- β 1 (59). This polymer was in paste form. The paste containing the growth factor TGF- β 1 was used to fill the grooves on the surface of a bioabsorbable fracture fixation pin. The self-reinforced pin was made of poly-LD-lactic acid. Prior to starting the study we made an in vitro assay in which sustained release of TGF- β 1 was demonstrated. A rat model was used to study the effect of TF- β 1 combined to the bioabsorbable

carrier. A rat distal femoral osteotomy was made and stabilized with the bioabsorbable fracture fixation pin containing 0, 5, or 50 μg of TGF- β 1. The study consisted of altogether 60 rats and the follow-up times were 1, 3, and 6 weeks. After sacrifice the femurs were examined radiographically, histologically, histomorphometrically, microradiographically, and with oxytetracycline labelling studies. We report enhanced bone formation around the osteotomy in the TGF- β 1-treated rats as compared to controls treated without the growth factor.

At the present, mostly preclinical, experimental data exists concerning the *in vivo* growth factor effects in clinically related animal models. An increasing number of positive results from experimental studies, such as ours, exist which could indicate a possible future success for growth factors within clinical orthopaedic practice. Nevertheless, it is not known whether or not the results from the animal studies can be transferred to humans with equal success. Recently, the first human studies using recombinant human BMP-2 (60,61,62) or BMP-7 (osteogenic protein-1, OP-1) (15,16,63,64) have been published. The results of these studies show a large variation among the responses of individual patients. These inconsistent results suggest that modulating factors known from animal studies affect the bone growth factor dependent bone induction process in humans as well. Such factors are the growth factor concentration, carrier properties and influence of other local and systemic growth factors and hormones. For better performance of bone growth factor containing bone-graft substitutes these factors need to be elucidated.

The use of bone growth factors will open a new set of treatment options. These will, ideally, enable us to obtain improved bone healing and bone formation in situations where the natural healing capacity of bone tissue is inadequate.

References

1. Lind M, Schumacher B, Søballe K, Keller J, Melsen F, Bünger C. Transforming growth factor- β enhances fracture healing in rabbit tibiae. *Acta Orthop Scand* 1993;64(5):553-556.
2. Nielsen HM, Andreassen TT, Ledet T, Oxlund H. Local injection of TGF- β 1 increases the strength of tibial fractures in the rat. *Acta Orthop Scand* 1994;65(1):37-41.
3. Beck LS, DeGuzman L, Lee WP, Xu Y, McFartridge LA, Gillett NA, Amento EP. Rapid publication. TGF beta-1 induces bone closure of skull defects:temporal dynamics of bone formation in defects exposed to rhTGF- β 1. *J Bone Miner Res* 1991;6(11):1257-1265.
4. Moxham JP, Kibblewhite DJ, Dvorak M, Perey B, Tencer AF, Bruce AG, Strong DM. TGF- β 1 forms functionally normal bone in a segmental sheep diaphyseal defect. *J Otolaryngol* 1996;25:388-392.
5. Canalis E, McCarthy TL, Centrella M. Growth factors and the regulation of bone remodeling. *J Clin Invest* 1988;81(2):277-281.
6. Solheim E. Growth factors in bone. *Int Orthop (SICOT)* 1998;22(6):410-416.
7. Groeneveld EHJ and Burger EH. Bone morphogenetic proteins in human bone regeneration. *Eur J Endocr* 2000;142(1):9-21.
8. Sandberg MM, Aro HT, Vuorio EI. Gene expression during bone repair. *Clin Orthop* 1993;289:292-312.
9. Aro HT, Kelly PJ, Lewallyn DG, Chao EYS. The effects of physiological dynamical compression on bone healing under external fixation. *Clin Orthop* 1990;256:260-263.
10. Neufeld G and Gospodarowicz D. Basic and acidic fibroblast growth factors interact with the same cell surface receptors. *J Biol Chem* 1986;261(12):5631-5637.
11. Ingber DE and Folkman J. Mechanochemical switching between growth and differentiation during fibroblast growth factor-stimulated angiogenesis in vitro: role of extracellular matrix. *J Cell Biol* 1989;109(1):317-330.
12. Hauscha PV, Mavrakos AE, Iafrati MD, Doleman SE, Klagsbrun M. Growth factors in bone matrix. *J Biol Chem* 1986;261(27):12665-12674.

13. Gombotz WR, Pankey SC, Bouchard LS, Phan DH, Puolakkainen PA. Stimulation of bone healing by transforming growth factor-beta 1 released from polymeric or ceramic implants. *J Appl Biomater* 1994;5(2):141-150.
14. Sherris DA, Murakami CS, Larrabee WFJr, Bruce AG. Mandibular reconstruction with transforming growth factor- β 1. *Laryngoscope* 1998;108(3):368-372.
15. Geesink RG, Hoefnagels NH, Bulstra SK. Osteogenic activity of OP-1 bone morphogenetic protein (BMP-7) in a human fibular defect. *J Bone Joint Surg Br.* 1999;81(4):710-718.
16. Groeneveld EH, van den Bergh JP, Holzmann P, ten Bruggengate CM, Tuinzing DB, Burger EH. Histomorphometrical analysis of bone formed in human maxillary sinus floor elevations grafted with OP-1 device, demineralized bone matrix or autogenous bone. Comparison with non-grafted sites in a series of case reports. *Clin Oral Implants Res* 1999;10(6):499-509.
17. Arnaud E, Morieux C, Wybier M, de Vernejoul MC. Potentiation of transforming growth factor- β 1 by natural coral and fibrin in a rabbit cranioplasty model. *Calcif Tiss Int* 1994;54(6):493-498.
18. Nixon AJ, Fortier LA, Williams J, Mohammed E. Enhanced repair of extensive articular defects by insulin-like growth factor-I-laden fibrin composites. *J Orthop Res* 1999;17(4):475-487.
19. Kawakami T, Uji H, Antoh M, Hasegava H, Kise T, Eda S. Squalene as a possible carrier for bone morphogenetic protein. *Biomaterials* 1993;14(8):575-577.
20. Kawakami T, Kawai T, Takei N, Kise T, Eda S, Urist MR. Evaluation of heterotopic bone formation induced by squalene and bone morphogenetic protein composite. *Clin Orthop* 1997;337:261-266.
21. Gao TJ, Lindholm TS, Marttinen A, Urist MR. Composites of bone morphogenetic protein (BMP) and type V collagen, coral derived coral hydroxyapatite, and tricalcium phosphate ceramics. *Int Orthop* 1996;20(5):321-325.
22. Sciadini MF, Dawson JM, Johnson KD. Evaluation of bovine-derived bone protein with a natural coral carrier as a bone graft substitute in a canine segmental model. *J Orthop Res* 1997;15(6):844-857.
23. Ohura K, Hamanishi C, Tanaka S, Matsuna N. Healing of segmental bone defects in rats induced by beta-TCP-MCPM cement combined with rhBMP-2. *J Biomed Mater Res* 1999;44(2):168-175.
24. Lee YM, Park YJ, Lee SJ, Ku Y, Han SB, Klokkevold PR, Chung CP. The bone regenerative effect of platelet-derived growth factor-BB delivered with chitosan/tricalcium phosphate sponge carrier. *J Periodontol* 2000;71(3):418-424.

25. Takaoka K, Nahara H, Yoshikawa H, Matsuhara K, Tsuda T, Ono K. Ectopic bone formation on and in porous hydroxyapatite combined with collagen and bone morphogenetic protein. *Clin Orthop* 1988;234:250-254.
26. Boden SD, Martin GJ, Morone MA, Ugbo JL, Moskovitz PA. Posterolateral lumbar intertransverse process spine arthrodesis with recombinant human bone morphogenetic protein 2/hydroxyapatite-tricalcium phosphate after laminectomy in the nonhuman primate. *Spine* 1999;24(12):1179-1185.
27. Rosenblum SF, Frenkel S, Ricci JR, Alexander H. Diffusion of fibroblast growth factor from a plaster of paris carrier. *J Appl Biomater* 1993;4(1):67-72.
28. Damien CJ and Parsons JR. Bone graft and bone graft substitutes: a review of current technology and applications. *J appl Biomater* 1991;2(3):187-208.
29. Nicoll SB, Rodan S; Santos EM, Tuan RS, Ducheyne P. In vitro release kinetics of biologically active transforming growth factor- β 1 from a novel porous glass carrier. *Biomaterials* 1997;18(12):853-859.
30. Nifuji A and Noda M. Local application of bone morphogenetic protein on developing chick embryos using fibrous glass matrix as a carrier. *Methods Mol Biol* 2000;135:465-469.
31. Lovell TP, Dawson EG, Nilsson OS, Urist MR. Augmentation of spinal fusion with bone morphogenetic protein in dogs. *Clin Orthop* 1989;243:266-274.
32. Park YJ, Ku Y, Chung CP, Lee SJ. Controlled release of platelet-derived growth factor from porous poly(L-lactide) membranes for guided tissue regeneration. *J Controlled Release* 1998;51(2-3):201-211.
33. Winn SR, Uludag H, Hollinger JO. Carrier systems for bone morphogenetic proteins. *Clin Orthop* 1999;367(Suppl):95-106.
34. Schmitz JP and Hollinger JO. A preliminary study of the osteogenic potential of a biodegradable alloplastic-osteoinductive alloimplant. *Clin Orthop* 1988;237:245-255.
35. Agrawal CM, Best J, Heckman JD, Boyan BD. Protein release kinetics of a biodegradable implant for bone non-unions. *Biomaterials* 1995;16(16):1255-1260.
36. Zellin G and Linde A. Importance of delivery systems for growth-stimulatory factors in combination with osteopromotive membranes. An experimental study using rtBMP-2 in rat mandibular defects. *J Biomed Mater Res* 1997;35(2):181-190.

37. Lucas PA, Laurencin C, Syftestad GT, Domb A, Goldberg VM, Caplan AI, Langer R. Ectopic induction of cartilage and bone by water-soluble proteins for bovine bone by using a polyanhydride delivery vehicle. *J Biomed Mater Res* 1990;24(7):901-911.
38. Pinholt EM, Solheim E, Bang G, Sudmann E. Bone induction by composite of bioerodible polyorthoester and demineralized bone matrix in rats. *Acta Orthop Scand* 1991;62(5):476-480.
39. Solheim E, Pinholt EM, Bang G, Sudmann E. Effect of local hemostatics on bone induction in rats: a comparative study of bone wax, fibrin-collagen paste, and bioerodible polyorthoester with and without gentamicin. *J Biomed Mater Res* 1992;26(6):791-800.
40. Buchholz RW, Henry SL, Henley MB. Fixation with bioabsorbable screws for the treatment of fractures of the ankle. *J Bone Joint Surg Am.* 1994;76(3):319-324.
41. Bergsma JE, deBrijn WC, Rozema FR, Bos RR, Boering G. Late degradation tissue response to poly(L-lactide) bone plates and screws. *Biomaterials* 1995;16(1):25-31.
42. Böstman OM, Pihlajamäki HK, Partio EK, Rokkanen PU. Clinical biocompatibility and degradation of polylevolactide screws in the ankle. *Clin Orthop* 1995;320:101-109.
43. Böstman OM. Osteoarthritis of the ankle joint after foreign-body reaction to absorbable pins and screws. *J Bone Joint Surg Br.* 1998;80(2):333-338.
44. Tielinen L, Manninen M, Puolakkainen P, Pihlajamäki H, Pohjonen T, Rautavuori J, Törmälä P. Polylactide pin with transforming growth factor- β 1 in delayed osteotomy fixation. *Clin Orthop* 1998;355:312-322.
45. Tielinen L, Manninen M, Puolakkainen P, Kellomäki M, Törmälä P, Rich J, Seppälä J, Rokkanen P. Inability of transforming growth factor-beta 1, combined with a bioabsorbable polymer paste, to promote healing of bone defects in the rat distal femur. *Arch Orthop Trauma Surg* 2001;121(4):191-196.
46. David SM, Gruber HE, Meyer RA, Murakami T, Tabor OB, Howard BA, Wozney JM, Hanley EN Jr. Lumbar spinal fusion using recombinant human bone morphogenetic protein in the canine. A comparison of three dosages and two carriers. *Spine* 1999;24(19):1973-1979.
47. Winn SR, Uludag H, Hollinger JO. Carrier systems for bone morphogenetic proteins. *Clin Orthop* 1999;367(Suppl):95-106.
48. Schmidmaier G, Wildemann B, Bail H, Lucke M, Stemberger A, Flyvbjerg A, Raschke M. Local liberation of IGF-I and TGF-beta 1 from a biodegradable poly(D,L-lactide) coating of implants accelerates fracture healing (Article in German). *Chirurg* 2000;71(9):1016-1022.

49. Vogelin E, Brekke JH, Jones NF. Heterotopic and orthotopic bone formation with a vascularized periosteal flap, a matrix and rh-BMP-2 (bone morphogenetic protein) in the rat model (Article in German). *Mund Kiefer Gesichtschir* 2000;4(Suppl 2):454-458.
50. Gombotz WR, Pankey SC, Bouchard LS, Ranchalis J, Puolakkainen P. Controlled release of TGF-beta 1 from a biodegradable matrix for bone regeneration. *J Biomater Sci Polym Ed* 1993;5(1-2):49-63.
51. Boyan BD, Lohmann CH, Somers A, Niederauer GG, Wozney JM, Dean DD, Carnes DL Jr, Schwartz Z. Potential of porous poly-D,L-lactide-co-glycolide particles as a carrier for recombinant human bone morphogenetic protein-2 during osteoinduction in vivo. *J Biomen Mater Res* 1999;46(1):51-59.
52. Isobe M, Yamazaki Y, Mori M, Amagasa T. Bone regeneration produced in rat femur defects by polymer capsules containing recombinant human bone morphogenetic protein-2. *J Oral Maxillofac Surg* 1999;57(6):695-698.
53. Lu L, Stamatias GN, Mikos AG. Controlled release of transforming growth factor beta 1 from biodegradable polymer microparticles. *J Biomed Mater Res* 2000;50(3):440-451.
54. Vert M, Chabot F, Leray J, Christel P. Stereoregular bioresorbable polyesters for orthopaedic surgery. *Makromol Chem* 1981;5(Suppl):30-41.
55. Leenslag JW, Pennings A, Bos RRM, Rozema FR, Boering G. Resorbable materials of poly(L-lactide) VII. In vivo and in vitro degradation. *Biomaterials* 1987;8(4):311-314.
56. Nakamura T, Hitomi S, Watanabe S, Shimizu Y, Jamshidi K, Hyon S-H, Ikada Y. Bioabsorption of polylactides with different molecular properties. *J Biomed Mater Res* 1989;23(10):1115-1130.
57. Kulkarni RK, Pani KC, Neuman C, Leonard F. Polylactic acid for surgical implants. *Arch Surg* 1966;93(5):839-843.
58. Hollinger JO and Battistone GC. Biodegradable bone repair materials. Synthetic polymers and ceramics. *Clin Orthop* 1986;207:290-305.
59. Tielinen L, Manninen M, Puolakkainen P, Päätiälä H, Pohjonen T, Rautavuori J, Rokkanen P. Combining transforming growth factor- β 1 to a bioabsorbable self-reinforced polylactide pin for osteotomy healing. An experimental study on rats. *J Orthop Sci* 1999;4:421-430.
60. Boyne PJ, Marx RE, Nevins M, Triplett G, Lazaro E, Lilli LC, Alder M, Nummikowski P. A feasibility study evaluating rhBMP-2/absorbable collagen sponge for maxillary sinus floor augmentation. *Int J Periodontics Restorative Dent* 1997;17(1):11-25.

61. Howell TH, Fiorellini J, Jones A, Alder M, Nummikoski P, Lazaro M, Lilly L, Cochran D. A feasibility study evaluating rh BMP-2/ absorbable collagen sponge device for local alveolar ridge preservation or augmentation. *Int J Peridontics Restorative Dent* 1997;17(2):124-139.
62. Cochran DL, Jones AA, Lilly LC, Fiorellini JP, Howell H. Evaluation of recombinant human bone morphogenetic protein-2 in oral applications including the use of endosseous implants:3-year results of a pilot study in humans. *J Peridontol* 2000;71(8):1241-1257.
63. Laursen M, Hoy K, Hansen ES, Gelineck J, Christensen FB, Bunger CE. Recombinant bone morphogenetic protein-7 as an intracorporal bone growth stimulator in unstable thoracolumbar burst fractures in humans:preliminary results. *Eur J Spine* 1999;8(6):485-490.
64. Friedlaender GE, Perry CR, Cole JD, Cook SD, Cierny G, Muschler GF, Zych GA, Calhoun JH, LaForte AJ, Yin S. et al. *J Bone Joint Surg Am.* 2001;83(Suppl 1)159-160.